Assessment of Physiological/Toxicological Effects of Camel Milk and Urine on Wistar Rats: A Study on Weight Gain, Haematological, Biochemical, and Histopathological Changes

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ABSTRACT

This experiment was steered to observe the safety evaluation of camel milk and its urine on the physiological indices of experimental rats. Four groups of twenty Wistar rats each were formed: Group 1 (Control), 2 milk (2 ml/100 g), 3 (2 ml/100 g), and 4 (milk/urine combination). The result showed that all treatment groups experienced a significantly decreased body weight gain across day 1-14, which significantly increased between day 18-21, except for group 4, which decreased significantly. The PVC, RBC, and haemoglobin increased non-significantly in group 2 and decreased in groups 3 and 4. WBC decreased significantly in milk/urine combination compared to other treatment groups. Neutrophils, Lymphocytes, and Monocytes did not show significant alteration across all the treatment groups. ALT decreased non-significantly (p>0.05) in all the treatment groups, while AST showed non-significant increased values in groups 2 and 3 except in group 4, which decreased significantly. Urea and creatinine decreased significantly (p<0.05) in all the treatment groups, while AST and ALT increased non-significantly (p<0.05) in group 4. NaCl showed significantly increased values across all the treatment groups while Bicarbonate decreased significantly in all the treatments. Ca2+ increased non-significantly. Histopathology results showed that group 4 has arrays of lesions compared to those treated separately. There was no observable lesion in these organs in group 2. It can be concluded that camel milk and its urine might have some beneficial effects when dosed separately, but they might predispose to harmful effects when combined.

INTRODUCTION

The World Health Organization estimated that the majority of the world’s population primarily uses medicines made from animals and plants (WHO, 2019), although reports have shown that animal products are more frequently used compared to plant resources (Glencross et al. 2020). In China, medicinal applications for around 1,500 animal species have been found (Deyrup et al., 2021). Around 250 animal species are supposedly utilised medicinally in North-East Brazil, whereas at least 109 animals have been employed for traditional medicinal purposes in India (Alves and Rosa 2005). A significant number of animals have reportedly been utilised in Ghana, South Africa, and Nigeria to treat various illnesses (Adeola, 1992; Mensah et al., 2019).

For millennia, camel milk and urine have been an essential part of traditional medicine and diets in numerous civilizations worldwide (Al-Moosawi et al., 2023). Modern scientific studies have confirmed the historical applications of camel milk and urine for medical purposes (Bakhsh et al., 2023).

Within traditional medicine, drinking camel urine either alone or when combined with milk is a well-known episode (Mok et al., 2021). This practice has been widely reported in Arabian nations, such as people of Bedouin communities who have a strong Islamic connection (Ali et al., 2019; Al-Yousef et al., 2012); among these tribes, the most highly prized urine is that of virgin female camels (Abdelzaher et al., 2020). But camel urine has historically...
been used more than other animal urine because it is much more alkaline (high potassium and magnesium levels, albuminous protein, and low percentages of uric acid, salt, and creatine) than other animal urine (Alkamees and Alsanad 2017).

Camel milk is vividly different from that of stomach animals (ruminant) milk in that it contains protective proteins such as lactoferrin, lactoperoxidase, immunoglobulin, and lysozyme, low cholesterol, low sugar, high minerals, and vitamin C (Yadav et al., 2015). Camel milk contains lactoferrin, which is ten times more abundant than lactoferrin in cow milk and has antimicrobial and antiviral properties like lysozyme and immunoglobulins (Swelum et al., 2021). According to reports, camel milk has low quantities of β-lactoglobulin and β-casein, making lactose intolerant experience hypersensitivity reactions (Konuspayeva et al., 2009).

Worries about possible health dangers of camel milk and urine and individuals sensitive to dairy products have been reported with significant adverse effects such as allergic reactions following its consumption (El-Aziz et al., 2022).

This research was designed to objectively access the physiological/toxicological indices in male Wistar rats.

Table 1: Twenty Male Wistar rats divided into 4 groups (n = 5), each consisting of five rats per group

<table>
<thead>
<tr>
<th>Grp</th>
<th>Description</th>
</tr>
</thead>
<tbody>
<tr>
<td>Grp 1</td>
<td>Control rats received normal saline orally for 21 days.</td>
</tr>
<tr>
<td>Grp 2</td>
<td>Received Camel’s milk at the dose of 2 ml/100 g by oral gavage for 21 days</td>
</tr>
<tr>
<td>Grp 3</td>
<td>Received Camel’s urine at the dose of 2 ml/100 g by oral gavage for 21 days</td>
</tr>
<tr>
<td>Grp 4</td>
<td>Received the combination of camel milk and its urine (1:1) at 1 ml/100 g by oral gavage for 21 days</td>
</tr>
</tbody>
</table>

Key: Grp = Group

Determination of weight gain

On days 1, 4, 7, 10, 13, 15, 18, and 21, the weights of the experimental mice were recorded using an automatic microelectronic scale (SDT™, London). A circular flexible bowl was put on the scale to measure the rat's weight, tarred to zero and subsequently measured.

Determination of haematological parameters

The entire blood in the EDTA bottles was utilized to assess various haematological parameters. PCV, HB Conc, and RBC were evaluated using Cole's method (Cole, 1986). Other parameters, including Total WBC, monocytes, lymphocytes, and neutrophils, were also assessed.

Determination of serum biochemical and electrolyte parameters

Creatinine, Blood Urea Nitrogen, Transferrase Alanine Phosphatase, and Aspartate Transferrase were analyzed. Serum electrolyte analysed includes sodium, potassium, calcium, chloride, and Bicarbonate following standard methods.

Animal sacrifice and tissues Sample Collection

The rats in all the groups were sacrificed humanely using ether as light anaesthesia. A small quantity of ether was placed on cotton wool to achieve this. The rat was placed in a small air-tight plastic container, and the ether-soaked cotton wool was placed on the nostril of the rat and subsequently covered. The anaesthetized rats were humanly killed after a few minutes. The heart, kidney, and liver were harvested for histopathology.

Histological procedures

The organs were appropriately treated with 10% formaldehyde (fixation) following the rats’ deaths to maintain their molecular makeup and structural integrity. The organs were dehydrated by gradually immersing these organs in a graded blend of ethanol and water. A solvent that could be mixed with the embedding media was used in place of ethanol. After injected xylene into the tissues, it cleared, or became transparent. The xylene-infused tissue was embedded in an oven with molten paraffin at 58 to 60 degrees Celsius. The solvent was able to evaporate due to the heat, and paraffin filled the spaces inside the tissues. After removing the tissue from the

https://scientifica.umyu.edu.ng/
oven, it solidified with its impregnating paraffin. Subsequently, the 5-micrometer pieces were submerged in water, moved to a glass slide, and stained with eosin and hemaphthalein stains. After the slides floated on the water, they were examined using a light microscope at a magnification of x 40 (Aremu et al., 2022).

Data analysis

Every created set of data was shown as mean ±Standard Deviation. Using the Dunnet Post-hoc multiple comparison test and the Graph Pad Prism statistical programme, the changes within the groups were examined using one-way ANOVA (www.Graphpad.com).

**Table 2: Changes in weight (g) gain of Wistar rats administered camel milk and its urine.**

<table>
<thead>
<tr>
<th>Days</th>
<th>Group 1</th>
<th>Group 2</th>
<th>Group 3</th>
<th>Group 4</th>
</tr>
</thead>
<tbody>
<tr>
<td>DAY 1</td>
<td>155.7±14.86</td>
<td>162.6±10.27</td>
<td>176.625±15.62</td>
<td>196.1±8.533</td>
</tr>
<tr>
<td>DAY 4</td>
<td>150.0±14.71</td>
<td>161.7±8.82</td>
<td>170.5±12.28</td>
<td>181.2±11.80</td>
</tr>
<tr>
<td>DAY 7</td>
<td>148.02±17.79</td>
<td>157.65±10.17</td>
<td>163.4±9.801</td>
<td>178.2±10.90</td>
</tr>
<tr>
<td>DAY 10</td>
<td>146.5±22.20</td>
<td>154.8±13.45</td>
<td>161.2±14.50</td>
<td>177.5±9.370</td>
</tr>
<tr>
<td>DAY 14</td>
<td>132.0±23.60</td>
<td>146.8±16.63</td>
<td>159.4±11.92</td>
<td>169.7±20.25</td>
</tr>
<tr>
<td>DAY 18</td>
<td>134.2±25.04</td>
<td>150.0±15.20</td>
<td>160.0±11.53</td>
<td>168.3±19.41</td>
</tr>
<tr>
<td>DAY 21</td>
<td>138.1±27.07</td>
<td>153.2±14.93</td>
<td>164.3±10.23</td>
<td>166.5±17.53</td>
</tr>
</tbody>
</table>

Values are expressed as mean ±SD

**Haematology following treatment with camel milk and its urine in Wistar rats**

The haematology following various treatments with camel milk and its urine showed that PVC, RBC, and haemoglobin (Hb) increased non-significantly in group 2 (camel milk) while it decreased significantly (P<0.05) in group 3 (camel urine) and 4 (milk and urine combination) when compared to group 1 (untreated control). MCV and MCHC relative reduction in camel milk and its urine groups while MCH relatively increased compared to the control group

**Table 3: Haematology of Wistar rats administered camel milk and its urine.**

<table>
<thead>
<tr>
<th>Parameters</th>
<th>Grp 1</th>
<th>Grp 2</th>
<th>Grp 3</th>
<th>Grp 4</th>
</tr>
</thead>
<tbody>
<tr>
<td>PCV (%)</td>
<td>38.50±2.08</td>
<td>39.25±2.50</td>
<td>35.80±3.56&lt;sup&gt;a&lt;/sup&gt;</td>
<td>35.25±3.59&lt;sup&gt;a&lt;/sup&gt;</td>
</tr>
<tr>
<td>RBC×10&lt;sup&gt;6&lt;/sup&gt;/µl</td>
<td>6.61±0.33</td>
<td>6.87±0.30</td>
<td>6.21±0.68</td>
<td>6.25±1.14</td>
</tr>
<tr>
<td>HB (g/dl)</td>
<td>12.22±1.05</td>
<td>12.91±0.95</td>
<td>11.30±1.34</td>
<td>10.71±1.12</td>
</tr>
<tr>
<td>MCV (fl)</td>
<td>58.28±5.08</td>
<td>57.18±4.26</td>
<td>57.42±5.29</td>
<td>58.23±4.93</td>
</tr>
<tr>
<td>MCH (pg)</td>
<td>18.48±1.76</td>
<td>18.80±1.64</td>
<td>17.80±4.26</td>
<td>17.80±1.79</td>
</tr>
<tr>
<td>MCHC (g/dl)</td>
<td>31.63±1.66</td>
<td>30.60±0.41</td>
<td>30.90±1.15</td>
<td>31.00±1.71</td>
</tr>
<tr>
<td>WBC×10&lt;sup&gt;3&lt;/sup&gt;/µl</td>
<td>7.35±2.08</td>
<td>7.90±1.64</td>
<td>7.10±1.04&lt;sup&gt;a&lt;/sup&gt;</td>
<td>6.39±1.04&lt;sup&gt;a&lt;/sup&gt;</td>
</tr>
<tr>
<td>Lymph (%)</td>
<td>65.25±3.86</td>
<td>63.31±4.14</td>
<td>64.20±5.26</td>
<td>66.50±1.73</td>
</tr>
<tr>
<td>Neutro (%)</td>
<td>31.50±3.11</td>
<td>35.50±3.78</td>
<td>33.20±4.82</td>
<td>30.00±1.63</td>
</tr>
<tr>
<td>Mono (%)</td>
<td>2.00±0.82</td>
<td>1.50±0.71</td>
<td>1.60±0.89</td>
<td>2.00±0.82</td>
</tr>
<tr>
<td>Platelet ×10&lt;sup&gt;3&lt;/sup&gt;/d/µl</td>
<td>1.97±0.82</td>
<td>2.03±0.81</td>
<td>2.08±1.13</td>
<td>1.84±0.44&lt;sup&gt;a&lt;/sup&gt;</td>
</tr>
</tbody>
</table>

Data represented as Mean ±SD; No. of Wistar rats in each group=5; Significant *P<0.05; Grp: Group

**Serum chemistry following treatment with camel milk and its urine in Wistar rats**

Serum chemistry following treatment with camel milk and its urine showed that ALT decreased non-significantly (p>0.05) in all the treatment groups, while AST showed non-significant increased values in groups 2 (camel milk) and 3 (camel urine) except group 4 (milk and urine) that decrease significantly when compared to the control. Urea and creatinine decreased significantly (p<0.05) when compared to the control group in each of the therapy groups (Table 4).

**RESULT**

**Effect of camel milk and its urine on body weight**

The weights of the rats reduced non-significantly (p<0.05) from day 1 to 14. The weight of the rats started increasing from day 18 to 21 except for group 4 (Camel milk and its urine combined), which showed a steady decrease in weight gain across all the treatment days. There is a significant change in the rats' weight compared to the untreated control.
Serum electrolytes following treatment with camel milk and its urine in Wistar rats

When compared to other treatment groups and the untreated control, the serum chloride ion of group 3 (camel urine) decreased significantly (p<0.05), but it increased significantly (p<0.05) in group 4 (milk/urine combination). Sodium ion (Na+) showed a significant (p<0.05) increase in values across all treatment groups when compared to the untreated control. Bicarbonate (HCO3-) decreased significantly in all the treatments compared to the control. Potassium (K2+) did not show any alteration in all treatments compared to the control, but calcium increased non-significantly in all treatments compared to the control (Table 4).

Table 4: Serum and Biochemical values of Wistar rats administered camel milk and its urine.

<table>
<thead>
<tr>
<th>PARAMETER</th>
<th>Grp 1</th>
<th>Grp 2</th>
<th>Grp 3</th>
<th>Grp 4</th>
</tr>
</thead>
<tbody>
<tr>
<td>ALT (mmol/l)</td>
<td>15.78±7.14</td>
<td>9.00±1.89a</td>
<td>13.58±2.70a</td>
<td>12.10±2.68a</td>
</tr>
<tr>
<td>AST (mmol/l)</td>
<td>31.03±9.08</td>
<td>30.93±14.75</td>
<td>30.66±25.25</td>
<td>26.63±11.32a</td>
</tr>
<tr>
<td>ALP (mmol/l)</td>
<td>23.63±11.41</td>
<td>17.15±6.99</td>
<td>15.20±7.55</td>
<td>17.23±3.62</td>
</tr>
<tr>
<td>Urea (mmol/l)</td>
<td>4.80±2.24</td>
<td>4.77±1.11</td>
<td>4.44±3.87</td>
<td>4.13±7.48</td>
</tr>
<tr>
<td>Creatinine. (µmol/l)</td>
<td>113.10±49.92</td>
<td>78.78±7.77</td>
<td>104.54±5.40</td>
<td>87.65±12.59</td>
</tr>
<tr>
<td>Potassium ion (µmol/l)</td>
<td>3.35±0.47</td>
<td>3.03±0.41</td>
<td>3.88±1.06</td>
<td>3.75±0.38</td>
</tr>
<tr>
<td>Chloride ion (µmol/l)</td>
<td>89.63±11.07</td>
<td>89.50±3.43</td>
<td>82.86±10.96a</td>
<td>96.28±14.26a</td>
</tr>
<tr>
<td>Sodium ion (µmol/l)</td>
<td>131.90±2.95</td>
<td>137.30±7.62</td>
<td>133.20±7.91</td>
<td>133.30±8.63b</td>
</tr>
<tr>
<td>Bicarbonate ion (µmol/l)</td>
<td>17.25±1.99</td>
<td>15.68±2.22</td>
<td>16.10±1.33a</td>
<td>17.03±4.58a</td>
</tr>
<tr>
<td>Calcium ion (µmol/l)</td>
<td>0.70±0.27</td>
<td>0.76±0.31</td>
<td>0.77±0.35</td>
<td>0.69±0.35</td>
</tr>
</tbody>
</table>

Data rep. as Mean ±SD: n=5
Significant *p≤0.05
Grp: Group

Histopathology result of the heart of Wistar rats administered camel milk and its urine.

A: Section of the heart of rats in the untreated control (H&E) showing no observable visible lesion
B: Section of the heart of rats treated with camel milk showing no observable visible lesion (H&E)
C: Section of the heart treated with camel urine with mild necrotizing myocarditis (arrows) (H&E)
D: The section of the heart of rats treated with camel milk and its urine showing severe necrotizing myocarditis (dotted circle) (H&E)
Histopathology result of the kidney of Wistar rats administered camel milk and its urine.

E) Section of the kidney of rats; the control showing no observable visible lesion (H&E).
F) Section of the kidney treated with camel milk without observable lesion (H&E)
G) Section of the kidney treated with camel urine showing patchy tubular epithelial coagulation necrosis (arrow) (H&E)
H) Section of the kidney treated with combination of camel milk and its urine with patchy tubular epithelial coagulation necrosis and peri-tubular inflammatory cells (Arrows) (H&E).

Histopathology result of the liver administered camel milk and its urine.

I) Section of the liver of rats; the control and untreated (H&E) showing no observable visible lesion
J) Liver section treated with camel milk showing moderate centrilobular cord atrophy (H&E).
K) Liver section treated with camel urine with moderate hepatocellular atrophy (H&E)
L) Liver section treated with a combination of camel milk and its urine showing severe necrotizing hepatitis and hepatocellular vacuolar degeneration (Doted circle) (H&E).
DISCUSSION

Reports have shown that significantly decreased body weight may be a pointer to various responses, such as treatment-induced toxicity (Aremu et al., 2023). The results from this study showed that the weight of the rats reduced from day 1-14 across all the treatment groups indicating a negative effect of the treatment on the growths of the rats. This result conforms with the report of a previous study by Al-Anazi et al. (2022), who recorded a significant weight reduction in rats fed with fermented camels’ milk and high-cholesterol diets. The initial weight loss in all groups might be due to the sudden introduction of camel milk and urine into the rat’s diets which might have disrupted their gut microbiome and digestion (Al-Beltagi et al., 2023). Another possible reason is that camel milk and urine are known to possess anti-inflammatory and anti-oxidative properties, which could have initially suppressed the growth of certain gut bacteria, leading to a temporary reduction in nutrient absorption and weight loss. The weight started increasing from day 18 to 21 across the treated groups. This might be attributed to the adaptation pathway to the nutritional profile of camel milk and urine that could have promoted the growth of beneficial gut bacteria over time, leading to improved nutrient absorption and weight gain. This assertion agrees with Ibrahim et al. (2018), who assessed the hepatocurative potentials of camel milk and urine on CCl\textsubscript{4} induced hepatotoxicity in albino rats showing altered weight gain across the duration of the experiment. The steady weight loss observed in group 4 (milk/urine combination) could have resulted in unexpected negative effects on nutrient absorption, leading to consistent weight loss.

According to reports, haematological indices are a key indicator of toxicity with a high degree of predictive ability for systemic toxicity when exposed to chemicals, medications, whole plants, or animal products (Aremu et al., 2023). This study suggests that camel milk and its urine have a deleterious effect on the haematological parameters, resulting in various degrees of anaemia as observed with decreased PCV, RBC, and haemoglobin values, especially in camel milk/urine combination. This outcome agrees with Manav et al. (2019), who reported significantly decreased PCV, RBC, and haemoglobin in rats treated with camel milk/urine combination. This study's result specifically showed a significantly decreased platelet count in group 4 (milk/urine combination), which can lead to blood clot abnormalities when taken for a long time.

The treated groups showed lower serum AST, ALT, and ALP activities, indicating that camel milk and/or urine have hepatoprotective activities. This observation agrees with Ibrahim et al. (2018), who stated that camel milk and its urine possess significant hepatoprotective potentials in tetrachloride-induced hepatotoxicity.

Serum electrolytes like urea and creatinine are usually used as major biomarkers in kidney function (Aremu et al., 2023). This present study showed no significant alterations in the kidney's level of creatinine, urea, Na, K\textsuperscript{+}, Cl\textsuperscript{-}, and HCO\textsubscript{3} in all the treatment groups. The creatinine and urea levels showed significantly decreased values, indicating that camel milk and urine are not nephrotoxic.

The histopathology results showed that the combination of camel milk and its urine (group 4) showed various degrees of lesion in the heart, kidneys, and liver compared to the other groups dosed separately. There was no observable lesion in these organs in group 2 (camel milk) compared to the untreated group.

The inference deduced from this work showed that camel milk/urine combination can pose a deleterious effect on these organs, leading to a marked toxicity when used for a long time.

CONCLUSION

Drawing on the study's findings, it can be said that camel milk and its urine have different effects on the body weight and health of rats. Camel milk and its urine might have some beneficial effects when dosed separately, but they might predispose to harmful effects when combined.

LIMITATIONS AND FURTHER RESEARCH

The scope of this research did not cover molecular and mechanistic approaches of assessing camel milk and urine toxicity in the biological system. Further studies could focus on evaluating the long-term effects of camel milk and urine exposure, considering different dosages and durations of treatment to assess both short-term and chronic impacts on animal health outcomes using molecular and mechanistic approaches.

ACKNOWLEDGMENT

We appreciate the technologist from the Department of Veterinary Pathology for the slides sectioning of all histopathology blocks.

CONFLICT OF INTEREST

No conflicting interest.

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