

ORIGINAL RESEARCH ARTICLE

Study on Adsorption of Alizarin Red S and Eriochrome Black T Dyes by Untreated Typha Grass as a Low Cost Adsorbent

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ABSTRACT

The purpose of this investigation was to study the adsorptive potential of untreated typha grass (UTG) using a batch system under control conditions to decolorize Alizarin Red S (ARS) and Eriochrome Black T (EBT) anionic toxic dyes from synthetic aqueous solution. In this regard, the impact of various experimental parameters namely, contact time, adsorbent dose, initial dye concentration, pH, and temperature were assessed and optimized. The adsorbent was characterized using Fourier transformed infrared (FTIR) spectroscopy, scanning electron microscopy (SEM), and point of zero charge (PZC) analysis. The maximum adsorption rates of ARS and EBT onto UTG were determined to be 49.92 mg/g and 47.43 mg/g, respectively, at equilibrium. In comparison to previous models evaluated, removal of both ARS and EBT by the UTG demonstrated that the data had a strong fit with pseudo-second-order kinetic and Freundlich adsorption isotherms. The contact time for both dyes was optimised at 15 minutes, while the pH for ARS and EBT were attained at 8 and 2 respectively. However, based on the established results of various parameters in this study, the adsorptive removal of ARS and EBT by UTG was a favourable process. The Van't Hoff plot was used to calculate the thermodynamic quantities of the adsorption process, including ΔG , ΔH and ΔS . The outcomes showed that the process was both spontaneous and thermodynamically possible. From the parameters tested, EBT was found to be better adsorbed onto UTG than ARS. Even though both dyes are anionic in nature, but EBT has additional diazo and nitro functional groups than the hydroxyl and sulphate functional groups present in ARS. This might be the reason for the better adsorption of EBT onto UTG than ARS. Results from this investigation supported the efficiency of UTG as a low-cost and environmentally friendly solution for the adsorption of ARS and EBT from contaminated wastewater.

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INTRODUCTION

The global discharge of industrial effluents into the environment poses substantial risks by polluting the ecosystem. This condition is a major cause of degradation of surface water and groundwater resources (Rashidi *et al.*, 2019; Rashidi *et al.*, 2021). One of the main sources of pollution which has a negative impact on aquatic life as well as human health is the discharge of hazardous coloured effluents from various manufacturing industries into aquatic media (Ghaderpoori and Dehghani, 2016). Contamination of the water bodies is one of the major pollution source involving majority of the industries discharging dyes, heavy metals, pharmaceutical waste, and various other organic pollutants (Khan *et al.*, 2018). Even at minute concentrations, these contaminants are non-biodegradable and can cause genetic and physiological issues in living organisms (Fu and Wang, 2011).

Dyes are complicated coloured aromatic compounds which are mostly of synthetic origins (Rashidi *et al.*,

2021). Synthetic dyes are widely used in the textile industries because they structurally feature reactive functional groups with high fiber-binding properties (Bilal and Asgher, 2015). It is estimated that 80,000 tons of dyes are produced and consumed almost yearly, making synthetic dyes one of the most damaging pollutants (Rossella *et al.*, 2021). Furthermore, it is evident that, human exposure to dyes may have serious health consequences, including allergies, cancer, mutagenesis, and teratogens (Gupta *et al.*, 2015).

Thus, environmentally, it has become desirable to treat wastewater containing dyes from industries (Rashidi *et al.*, 2021). The adsorption method especially at low concentrations, is the most effective and economical way to treat wastewater containing dyes (Putro *et al.*, 2021). This method however depends on the adsorption capacity of the adsorbent under consideration (Putro *et al.* 2021).

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This study aims to examine the adsorptive ability of untreated (raw) typha grass (UTG) by utilizing a batch adsorption approach for the decolorization of anionic dyes, ARS and EBT from wastewater. This grass was chosen because of its availability, affordability, and environmental friendliness (Taufiq *et al.*, 2018).

MATERIALS AND METHODS

General

The stock of chemicals used in this research were obtained from Sigma-Aldrich Ltd., London. These includes: Hydrochloric acid, sodium hydroxide, Alizarin Red S (ARS) dye and Eriochrome Black T (EBT) dye. The chemicals were used without further purification, and all solutions were prepared with distilled water. The scanning electron microscopy images were recorded using Phenom World Eindhoven microscope, while the Fourier Transformed Infrared Spectroscopy was recorded on Cary 630 FTIR Spectrophotometer from

Agilent Technology. UV-Visible spectroscopy was performed on Perkin-Elmer Lambda 35 UV-Visible spectrometer.

Preparation of Dye Solutions

To make 1000 mg/L of the stock solution for each of the commercial grade Alizarin red S and Eriochrome black T dyes, 1g of each ARS and EBT dyes were weighed and dissolved differently into 1L volumetric flask and made up to the mark using distilled water. The stock solution was diluted with distilled water to prepare the experimental solutions at the desired concentration. Subsequently, the concentrations of the dyes solutions were measured using UV-Visible spectrometer (Perkin Elmer, Lambda 35) at maximum wavelengths of 424.89 nm and 524.44 nm for ARS and EBT, respectively. The chemical structure of the studied dyes is as presented in Figure 1.

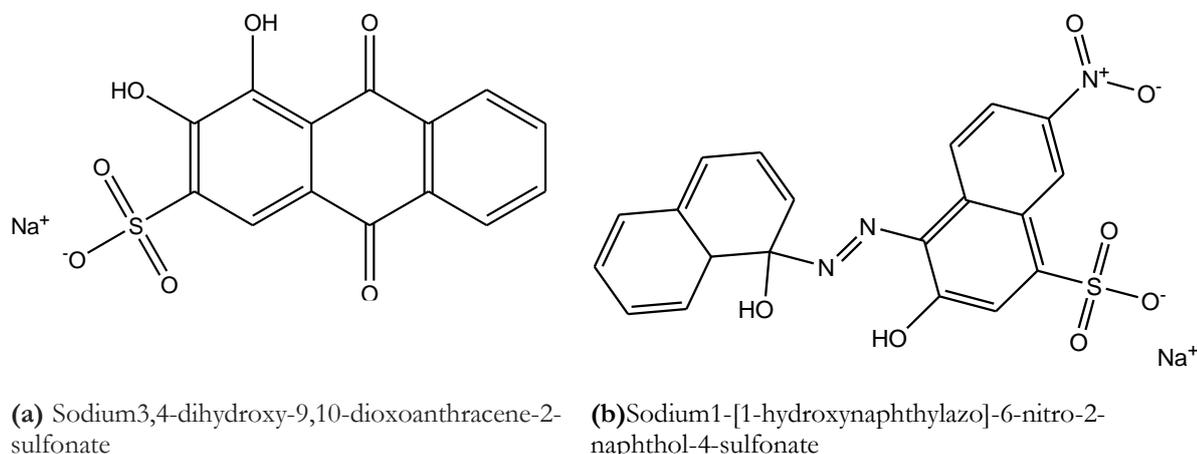


Figure 1: Chemical Structures of (a) Alizarin Red S and (b) Eriochrome Black T Dyes

Preparation and Characterization of Typha Grass (UTG)

The typha grass was obtained from Marma town, Guri local government area of Jigawa State, Nigeria. The sample was first washed with tap water then distilled water to remove the impurities (Nwosu *et al.*, 2017). The sample was air-dried and then oven dried at 105°C. The dried sample was untreated, pulverized and passed through a 2mm sieve.

The physical properties of the adsorbent, such as moisture content, bulk density, and pore volume, were determined using the methods reported by Ali *et al.* (2018); Ayuba and Idoko (2020). As reported by Bakatula (2018), the point of zero charge (PZC) of UTG was investigated using the salt addition method. The surface morphology before/after adsorption were studied using scanning electron microscopy (SEM). While the Fourier transformed infrared (FTIR) spectroscopy was also recorded both before and after dye adsorption.

Batch Adsorption Study

To find the ideal conditions for the equilibrium adsorption of ARS and EBT onto UTG, batch experiments were conducted and the optimized condition was found. Thus, in a 250 cm³ conical flask, each system was operated independently at the temperature of 30°C, 40°C, and 50°C. The conical flasks were covered during the equilibration period and set up on an Innova 4000 incubator shaker with temperature control for the optimal times. The substance was filtered through Whatman No 1 filter paper after attaining adsorption equilibrium. The filtrate was examined using UV-Visible spectrometer. Then the efficiency of the dyes to adsorb from their aqueous solutions onto UTG was calculated using equations 1 and 2 as reported by Bansal *et al.* (2020) and Ayuba and Idoko (2021).

$$\text{Adsorption Capacity (qe)} = \frac{(C_0 - C_e) \times V}{m} \quad (1)$$

$$\% \text{ Adsorption} = \left(\frac{C_0 - C_e}{C_0} \right) \times 100 \quad (2)$$

Where V is the volume of dye solution (L), m is the mass (g) of the adsorbent, and q_e is the adsorption capacity (mg/g). C_0 and C_e are the initial and final equilibrium concentrations (mg/l) of the dyes in solution.

RESULTS AND DISCUSSION

Adsorbent Characterization

The physical characteristics of the adsorbent, including bulk density (organic matter), moisture content, and pore volume, are presented in Table 1. The results showed that the adsorbent exhibited good adsorptive properties for removing anionic dyes from aqueous solution owing to its low moisture content, high pore volume, and bulk density. Ayuba and Thomas (2021) reported similar results when they used carbonized Bambara groundnut (*Vigna subterranean*) shells to remove Paraquat dichloride from aqueous solution.

Table 1: Physical properties of untreated typha grass (UTG)

Moisture Content (%)	Bulk Density (gcm^{-3})	Pore Volume (cm^3)
17.80	0.397	1.253

Figure 2 shows the graph of the point of zero charge (PZC) for UTG adsorbent. Since there was no surface charge at the PZC that could be neutralized by ions at the adsorbent's surface, any adsorbed ions that do exist must be bonded in surface complexes. The pH of the neutral medium, or the region where the equilibrated ions in an aqueous solution are, is around 7, which indicates the presence of complete charge balance in the UTG under study. Moreover, substrates with low PZC values have the strongest tendency to treat effluents contaminated with cations, although the substrates with high PZC values would be better suited to adsorb anions (Nasiruddin and Anila, 2007; Zhang *et al.*, 2020).

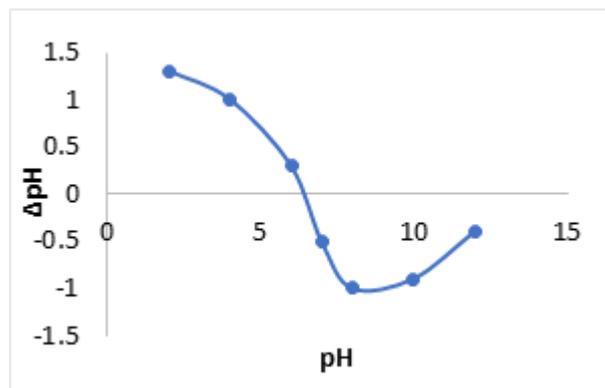


Figure 2: Measurement of point of zero charge (PZC) of UTG

The study of an adsorbent's surface morphology is essential in determining how well-suited it is to interact with the species it comes into contact with (Bansal *et al.*, 2020). Figure 3 gives the result of surface morphology analysis of UTG characterized by SEM methodology. SEM was employed to identify any surface morphological changes that might occur on UTG prior to and after the adsorption of ARS and EBT dyes. The instrument was operated at x500 magnification and a 15.00 kV accelerating voltage (Ali *et al.*, 2018). Whereas figure 3a indicates that the surface of UTG was rough, more porous, and more amorphous before the adsorption of each dye than it was after. In Figure 3b and c, earlier pores and rough surfaces in the un-adsorbed UTG have significantly decreased in comparison to Figure 3a because of the dyes' adsorption onto its surface. Secondly, it is obvious from comparing Figures 3b and c that EBT covers a greater proportion of UTG than ARS. Other authors have reported these observations as an indicator and evidence of adsorption of species onto adsorbents (Madhushika *et al.*, 2020).

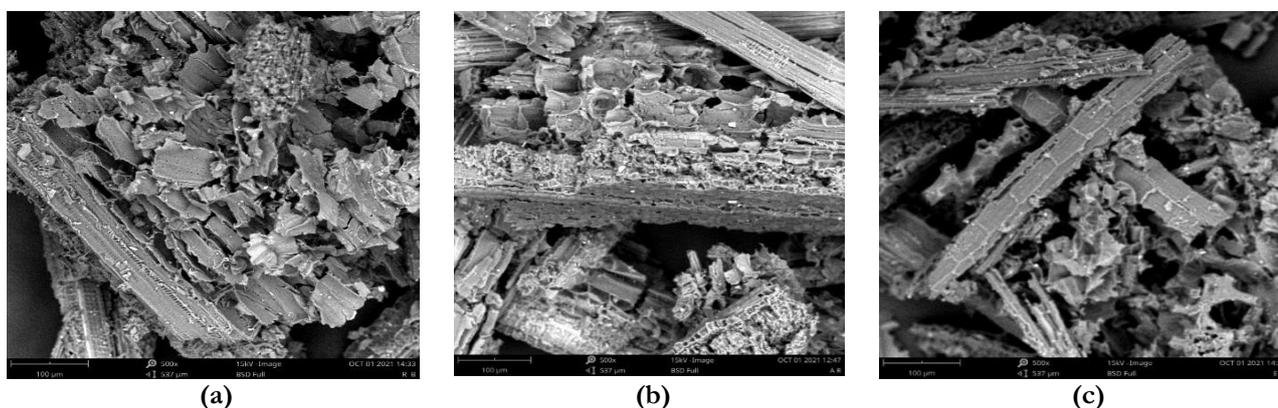


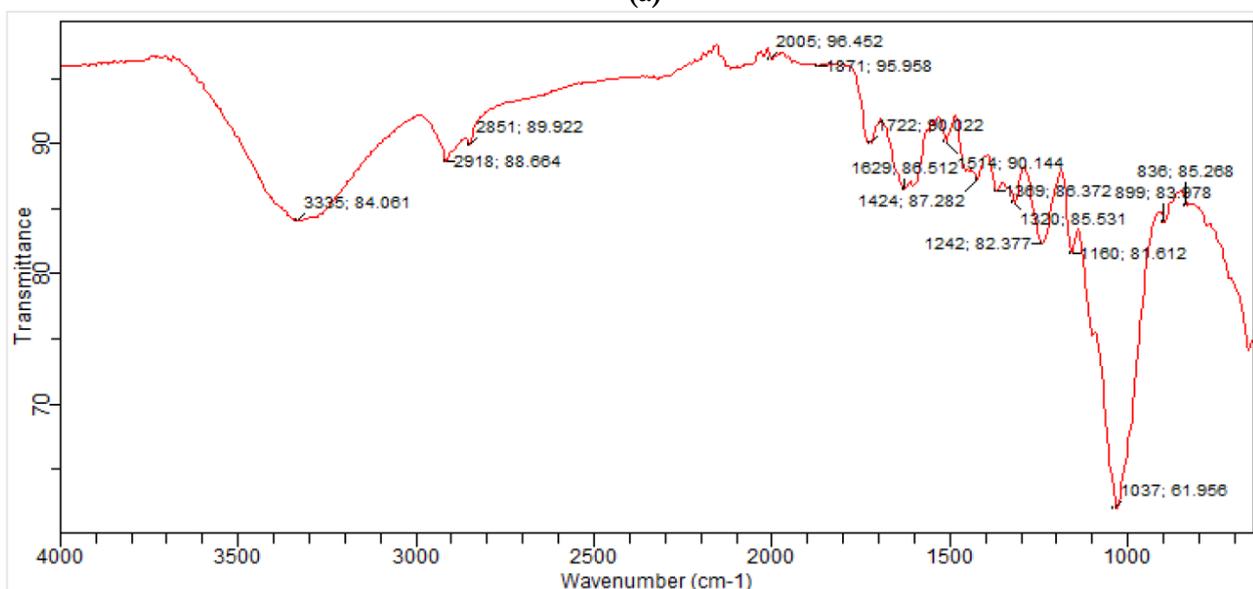
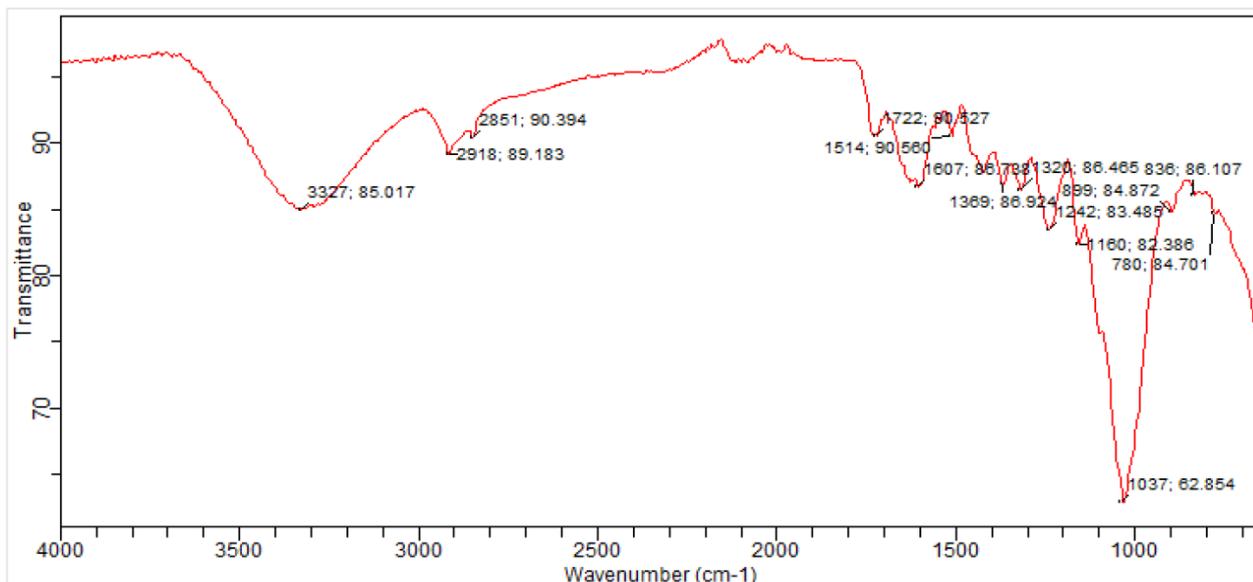
Figure 3: SEM micrographs of (a) UTG before adsorption and (b) after adsorption onto ARS and (c) after adsorption onto EBT.

FTIR was used to identify the functional groups that were present in the samples. Figures 4a–c illustrates the

FTIR spectra of UTG both before and after ARS and EBT adsorption. The significant distinctive peaks

identified at 3327 cm^{-1} , 2918 cm^{-1} , 1607 cm^{-1} , 1514 cm^{-1} , and 1104 cm^{-1} before adsorption were connected to the stretching vibration mode of O-H, C-H, C=O, C \equiv C, and C-O groups, respectively (Ali, *et al.*, 2018). After loading UTG with ARS and EBT, there was a notable change in the vibration frequency of the free OH, C=O, and C-O stretches. Possible causes of the vibration frequency changes include the binding of ARS and EBT ions to the surface of the adsorbents (Babalola *et al.*, 2016). Because some of the important adsorption sites characteristics of effective adsorbents comprise hydroxyl, carbonyl, and

carboxylic acid (Itodo *et al.*, 2011). The variation in absorption bands of the detected functional groups before and after adsorption were reported in Table 2. Different absorption bands were not witnessed before and after adsorption of the dyes onto UTG in C-H and C \equiv C stretches, an indication that they did not partake in the adsorption process. But those in O-H (for ARS) and C-O (for EBT) and C=O stretch (for both ARS and EBT) are indicators of the functional group likely used for the adsorption of the dyes onto UTG.



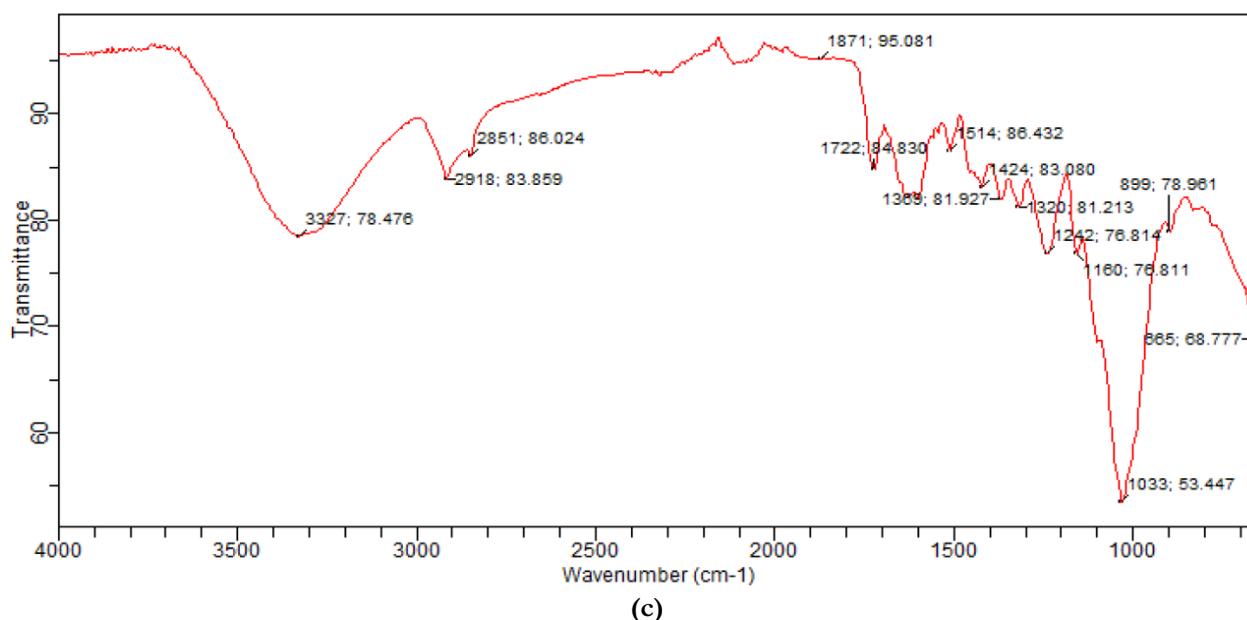


Figure 4: FTIR spectra of UTG (a) before adsorption (b) after ARS adsorption (c) after EBT adsorption.

Table 2: Summary of FTIR absorption bands of UTG before and after adsorption of ARS and EBT

Functional Group	Absorption range (cm ⁻¹)	Before absorption (cm ⁻¹)	After absorption onto ARS (cm ⁻¹)	Difference (cm ⁻¹)	After absorption onto EBT (cm ⁻¹)	Difference (cm ⁻¹)
OH stretch	3700-2500	3327	3335	13	3327	0
C-H stretch	3000-2840	2918	2918	0	2918	0
C=O stretch	1710-1580	1607	1629	12	1722	115
C≡C stretch	675-1550	1514	1514	0	1514	0
C-O stretch	1080-1300	1104	1104	0	1160	56

Batch Adsorption Studies

Effect of Contact Time

The influence of the contact time between 0-120 min of ARS and EBT removal by UTG was investigated in a suspension of 50 mg/L with 0.1g adsorbent at neutral pH. Adsorption happened rapidly at the beginning of the process until the equilibrium was reached within the first 10min for EBT and 15min for ARS (Figure 5a). During the duration, up to a maximum of 120 minutes, adsorption approached a constant condition. This optimum adsorption contact times were used for each of the dyes during batch adsorption studies. The abundance of unoccupied sites on an adsorbent's surface is connected to the rapid adsorption of dyes by the adsorbent at the beginning of contact time (Rashidi *et al.*, 2021). As the time progresses to a maximum, the unoccupied sites would then be filled with dye molecules, eventually leading to desorption (Nourmoradi *et al.*, 2012).

Effect of Adsorbent Dosage

The impact of the adsorbent dose (g) on the ARS and EBT dyes was investigated by adsorption experiments. The adsorption was conducted at neutral pH, room

temperature, optimal contact time for each dye, dosage range of 0.02g-0.2g, initial dye concentration of 50 mg/L. The adsorption capacity of ARS and EBT decreased as the adsorbent dose increased within the range of 0.02g-0.2g, attaining results of 49.92 mgg⁻¹ and 47.43 mgg⁻¹ for ARS and EBT, respectively (Figure 5b). Therefore, the highest adsorption capacity was obtained at a dosage of 0.02g of the adsorbent for both dyes. However, this dosage was chosen as the optimum rate for subsequent batch adsorption experiments. The availability of unoccupied active sites on the surface of the adsorbent was determined to be the main reason why the adsorption capacity of ARS and EBT increased. In comparison to substantially greater doses, the lowest dose (0.02g) was found to be more effective at adsorbing ARS and EBT (Bansal *et al.*, 2020). However, it was discovered in literature that as adsorbent dose increases, the effectiveness of the removal of dyes significantly improves (Li *et al.*, 2019). This behaviour is related to the large availability of empty adsorption sites during the initial stages of adsorption, followed by an increase in the repulsive forces brought on by the presence of the

adsorbed ions, making it more challenging to access the remaining sites (Nwosu *et al.*, 2017).

Effect of Initial Dye Concentration

The effect of the initial dye concentration in the range of 20-120 mg/L on the dye adsorption was investigated at room temperature and the contact time and adsorbent dosage were optimized. Figure 5c demonstrates a considerable increase in ARS and EBT adsorption with increasing initial dye concentration. The presence of more molecular ions in the solution, which compete with the available binding sites on the surface of the adsorbent, is responsible for this considerable change in the adsorption capacity of UTG with a rise in both ARS and EBT concentrations (Pahalagedara *et al.*, 2014). However, increase in ARS and EBT concentration undesirably affects the removal of the dyes. While the removal of ARS and EBT decreased as the initial dye concentration was increased from 20 to 120 mg/L (Liu *et al.*, 2015). The electrostatic attraction between the dye molecules in solution and the dye molecules adsorbed by

the UTG is increased as the initial dye concentration increases.

Effect of pH

By varying the solution pH values from 2 to 10, the influence of pH on the adsorption of ARS and EBT onto the surface of UTG was evaluated. Figure 5d demonstrated that as the pH of the ARS solution increased, the adsorption capacity of UTG decreased as the pH of the EBT solution increased (Zubair *et al.*, 2017). Therefore, at pH levels of 8 and 2, respectively, the optimum adsorption capacity for ARS was 47.99 mg/g while for EBT was 49.44 mg/g. This outcome could be explained by variations in the adsorbent's surface charge. However, by lowering the pH of the solution, the amount of positive charges on an adsorbent's surface improves adsorption (Rashidi *et al.*, 2021); in the same way, it increased ARS's ability to adsorb in more acidic pH solutions. In other words, for EBT, the electrostatic repulsions between the negatively charged adsorbent and anionic dyes increased at a basic pH (Raveendra *et al.*, 2015).

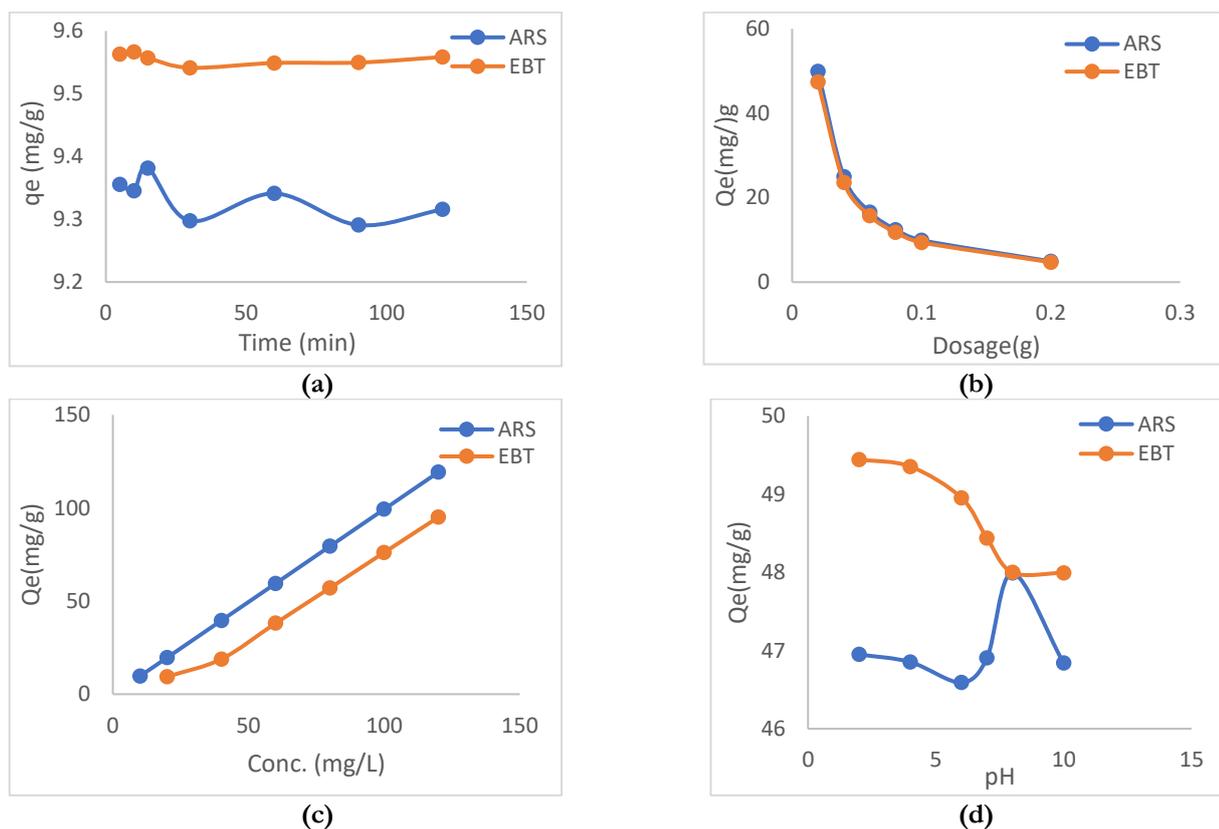


Figure 5. Effect of (a) contact time, (b) dosage, (c) initial dye concentration, (d) pH, on the adsorption of ARS and EBT onto UTG

Adsorption Isotherms

The adsorption isotherms are useful for determining how the adsorbate is distributed on the adsorbent's surface under equilibrium conditions (Sharma *et al.*, 2018). To identify the mechanistic characteristics of interactions between the adsorbent and the adsorbate at equilibrium, Langmuir, Freundlich, Temkin, and D-R models were

tested (Bansal *et al.*, 2020). The models were employed to evaluate the removal of ARS and EBT by UTG. The linearized form of these models are presented in equations (3-6) as reported by Sadani *et al.* (2020).

$$\frac{1}{q_e} = \frac{1}{Q_0} + \frac{1}{Q_0 K_L C_e} \quad \text{Langmuir} \quad (3)$$

$$\text{Log}Q_e = \text{log}K_f + \frac{1}{n} \text{log}C_e \quad \text{Freundlich} \quad (4)$$

$$q_e = B_T \ln C_e + B_T \ln K_T \quad \text{Temkin} \quad (5)$$

$$\ln q_e = \ln q_m - \beta \epsilon^2 \quad \text{Dubinin Radushkevich (D-R)} \quad (6)$$

Where C_e stands for the equilibrium dye concentration (mg/L); q_e and Q_0 for the equilibrium and maximum adsorption capacities (mg/g); and K_L , K_F , K_T , and β for the Langmuir, Freundlich, Temkin, and D-R constants, respectively. The intensity of the bond and the value of n indicate that ARS and EBT can be successfully adsorbed onto UTG (Nourmoradi *et al.*, 2015). Table 3 displays the calculated values of the isotherm parameters for ARS and EBT removal by UTG.

Table 3: Adsorption Isotherm constants for the adsorption of ARS and EBT onto UTG

Parameter	Alizarin Red S	Eriochrome Black T
Langmuir		
$Q_0(\text{mg/g})$	-9.3720	-151.51
$K_L(\text{L/mg})$	-0.1680	-0.091
R_L	-0.1115	-0.222
R^2	0.967	0.994
Freundlich		
$1/n$	1.0029	1.1421
n	0.4992	0.8755
K_F	1.2083	16.270
R^2	0.958	0.990
Temkin		
A_T	0.5288	1.445
b_T	86.09	53.415
B	29.259	47.161
R^2	0.827	0.92
Dubinin-Radushkevich		
$q_m(\text{mg/g})$	48.638	85.567
$K_{dr}(\text{mol}^2/\text{KJ}^2)$	3.0×10^{-6}	4.0×10^{-6}
$E(\text{kJ/mol})$	4.0×10^{-1}	1.12×10^{10}
R^2	0.841	0.901

However, from the results obtained, the Freundlich isotherm established a greater correlation coefficient (R^2) value than Langmuir, Temkin and D-R isotherms. Thus, it can be said that the Freundlich isotherm model provided the greatest match for the experimental data of ARS and EBT removal by the UTG (Zhou *et al.*, 2018).

Table 4: Kinetic models for the adsorption of ARS and EBT onto UTG

Pseudo-first order	Parameter	$q_{e,Cal}(\text{mg/g})$	$q_{e,Exp}(\text{mg/g})$	$K_1(\text{min}^{-1})$	R^2
	ARS	46.682	0.328	0.019	0.120
	EBT	49.511	0.124	0.006	0.004
Pseudo-second order	Parameter	$q_{e,Cal}(\text{mg/g})$	$q_{e,Exp}(\text{mg/g})$	$K_2(\text{min}^{-1})$	R^2
	ARS	46.682	46.511	0.770	0.999
	EBT	49.511	49.504	0.020	0.998

This suggests that the physical adsorption process is in control of the adsorption of ARS and EBT dyes using UTG as an adsorbent and validates the existence of heterogeneous adsorption sites on UTG (Ahmed *et al.*, 2021). Furthermore, the obtained result revealed that the removal of ARS and EBT by a variety of adsorbents, including modified kaolinite clay, NiFe_2O_4 magnetic nanoparticles, and Mosambi peel activated carbon, follows a similar isotherm model as reported by Zong *et al.*, (2016) and Sharma *et al.*, (2018).

Kinetic Studies

Data from batch adsorption studies were tested against four kinetic models in order to understand the adsorption kinetics of ARS and EBT dyes onto UTG. These includes kinetic explanations of the adsorption of these dyes onto UTG adsorbent using pseudo-first-order, pseudo-second-order, Elovich, and intraparticle diffusion (Alinejad *et al.*, 2019). The kinetics models employed have a linear form as shown in equations (7–10) as reported by Ayuba *et al.* (2020) and Kamarehie *et al.* (2020).

$$\log(q_e - q_t) = \log(q_e) - \frac{k_1}{2.303} t \quad (\text{Pseudo - First order}) \quad (7)$$

$$\frac{t}{q_t} = \frac{1}{k_2 q_e^2} + \frac{1}{q_e} (t) \quad (\text{Pseudo - Second order}) \quad (8)$$

$$q_t = 1/\beta \ln(\alpha\beta) + (1/\beta) \ln t \quad (\text{Elovich}) \quad (9)$$

$$q_t = C + K_{int} t^{1/2} \quad (\text{Intraparticle Diffusion}) \quad (10)$$

Where q_e and q_t stand for the quantity of adsorption at equilibrium and at the time, t , and k_1 , k_2 and k_{int} stand for the pseudo-first order, pseudo-second order, Elovich and Intraparticle diffusion rate constants, respectively. However, Table 4 gives the kinetics of the ARS and EBT adsorption onto UTG as well as the values of the rate constants and q_e as determined by the pseudo-first, pseudo-second order, Elovich and Intraparticle diffusion. Furthermore, the results from the aforementioned models showed that, in comparison to other models considered, the correlation coefficient value (R^2) favoured by pseudo-second order. Additionally, the pseudo second order model kinetics used to describe the ARS and EBT adsorption by UTG can also clearly be seen when comparing the $q_{e,Cal}$ and $q_{e,Exp}$ of the tested models (Li *et al.*, 2019; Yao *et al.*, 2020).

Table 4: Continued

Elovich	Parameter	B	A	R ²
	ARS	178.571	1.2×10 ⁸¹	0.039
	EBT	68.493	4.1×10 ¹⁷¹	0.011
Intraparticle diffusion	Parameter	K ₃	C	R ²
	ARS	0.004	46.508	0.142
	EBT	-0.015	12.02	0.001

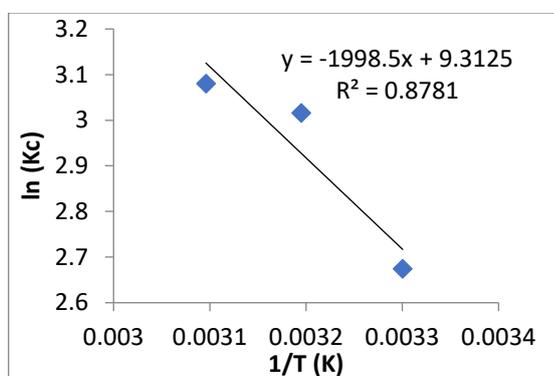
Thermodynamic Studies

Thermodynamic studies provide vital information on the energetics associated with the adsorption process. The thermodynamics of an adsorption process can be explained using the changes in enthalpy (ΔH), entropy (ΔS) and Gibb's free energy (ΔG). Three different temperatures (298K, 308K and 318K) were used for the batch adsorption studies. Equation (11) was used to determine how the Gibb's free energy changed for the adsorption. Equation (12) was used to calculate the

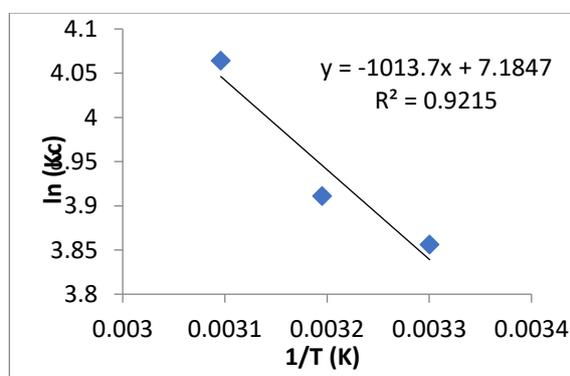
changes in entropy and enthalpy by plotting lnK_L against 1/T to give Van't Hoff's plot (Figure 6). ΔH was calculated using the graph's slope, whereas ΔS was calculated using the plot's intercept (Min *et al.*, 2012; Umar *et al.*, 2021).

$$\Delta G = -RT \ln K_L \tag{11}$$

$$\ln K_L = \frac{\Delta H}{RT} + \frac{\Delta S}{R} \tag{12}$$



(a)



(b)

Figure 6: Van't Hoff plot for (a) ARS and (b) EBT adsorption at different temperatures

Summary of the results obtained from the thermodynamic evaluation are as presented in table 5. The adsorption system was thermodynamically feasible and spontaneous due to the negative values of ΔG, and with values lower than -20KJ/mol, the adsorption process for both dyes was physical in nature (Nourmoradi *et al.*, 2015; Ayuba and Idoko, 2020). As the temperature increases from 298 to 318K, the rate of adsorption increased. Furthermore, the endothermic nature of the adsorption of ARS and EBT by UTG was

confirmed by the positive value of enthalpy change ΔH. In addition, the results showed that the ΔH values were positive and lower than 80 kJ/mol, indicating that the adsorbate and adsorbent surface were physically interacting during the adsorption process. Moreover, positive entropy change (ΔS) values indicated that UTG can easily loose off ARS and EBT and that the adsorption phenomena is entropy-controlled rather than enthalpy-controlled (Asadullah *et al.*, 2019).

Table 5: Thermodynamic parameters for the adsorption of ARS and EBT onto UTG

Dye	T (K)	K _c	ΔG (KJ/mol)	ΔH (KJ/mol)	ΔS (J/mol.K)
ARS	303	14.503	-6.737	16.616	77.424
	313	20.399	-7.847		
	323	21.757	-8.271		
EBT	303	65.439	-10.177	8.283	53.819
	313	49.018	-10.532		
	323	98.901	-12.337		

CONCLUSION

In this study, anionic dyes ARS and EBT were extracted from aqueous solution using a batch method, and UTG was employed as a cheap, eco-friendly and cost-effective adsorbent. Some physical parameters of the adsorbent were analysed and found the adsorbent to be promising. The surface of the adsorbent was characterized in order to assess the extent of interaction between studied dyes and the surface. The batch system performance was optimized based on how the contact time, adsorbent dose, initial dye concentration, temperature and pH affect adsorption efficiency. The pseudo-second-order kinetic and Freundlich isotherm models performed well in describing the experimental data for both dyes, according to the results. The adsorption process was thermodynamically feasible, spontaneous and adhered to the physical adsorption of dyes mechanism. Maximum adsorption capacity obtained for ARS and EBT were 47.99 mg/g and 49.44 respectively. This is an indication that EBT was better adsorbed than ARS even though they were both anionic in nature. Presence of diazo and nitro functional groups in EBT in addition to hydroxyl and sulphate functional groups present in ARS may be the reason for the better adsorption of EBT onto UTG over ARS. The findings showed how successfully UTG may be utilized to remove anionic dyes from aqueous medium. Specifically, the findings suggested that UTG might prove to be a useful adsorbent for the treatment of ARS and EBT-contaminated water.

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CONFLICT OF INTEREST

The authors state that they are free of any conflicts of interest that might have affected the work presented for the current study.

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